

Roles of H₂O₂ and OH• radical in bactericidal action of immobilized TiO₂ thin-film reactor: An ESR study

Guoliang Yan^a, Jian Chen^b, Zhaozhe Hua^{b,*}

^a College of Food Science and Nutrition Engineering, China Agriculture University, Beijing 100083, PR China

^b Key Laboratory of Industrial Biotechnology, Ministry of Education, Jiangnan University, 1800 Lihu Road, Wuxi 214122, PR China

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ABSTRACT

To evaluate the bactericidal roles of hydrogen peroxide (H₂O₂) and hydroxyl radical (OH•) in immobilized titanium dioxide (TiO₂) thin-film reactor, the mechanism of H₂O₂ and OH• formation in photocatalytic reaction was investigated by applying electron spin resonance (ESR) technique. The results showed that the formation of OH• largely depended on H₂O₂ formed in photocatalytic reaction. OH• were mainly originated from the H₂O₂ via the reduction reaction although a relatively small portion came from the direct photolysis of H₂O₂. Due to the convertible relationship between H₂O₂ and OH•, the approaches that can increase H₂O₂ concentration such as bubbling air or adding H₂O₂ could greatly result in enhancement of OH•. The results of bactericidal experiments showed that killing effect was completely inhibited by catalase, in contrast, the inactivation efficiency was proportionally increased with elevated H₂O₂ levels. Considering the fact that H₂O₂ alone was not enough to complete the disinfection action, it is concluded that OH•, not H₂O₂ was the primary bactericidal radicals in the immobilized TiO₂ thin-film reactor; however, due to its conversion to OH• and oxidative toxicity, H₂O₂ was also indispensable for photocatalytic bactericidal effect.

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1. Introduction

In 1985, Matsunaga et al. reported for the first time the antibacterial effect of TiO₂ photocatalytic reactions [1]. Since then, the biological effect of TiO₂ photocatalyst has attracted increasing attention worldwide because of its stable and strong oxidative reactions resulting in biocidal activity. It is generally accepted that the mechanism of cell death by the TiO₂ photochemical reaction involves photogenerated holes and reactive oxygen species (ROS) such as superoxide radical (O₂⁻), hydrogen peroxide (H₂O₂) and hydroxyl radical (OH•). Mechanisms for the primary events occurring at the catalyst surface have been described [2,3]. Upon excitation by light whose wavelength is less than 385 nm, an electron (e⁻) is promoted from the valence band to the conduction band of the semiconducting oxide leaving a positive hole (h⁺) in the valence band. The positive hole can oxidize H₂O to OH• (Eqs. (1) and (2)), and the negative electron can reduce oxygen to O₂⁻ or H₂O₂ in the presence of molecular oxygen (Eqs. (3) and (4)).



The photogenerated H₂O₂ and O₂⁻ will then promote the generation of OH• in TiO₂ photocatalytic reactor (Eqs. (5) and (6)).



In addition, OH• can be formed from the photolytic decomposition of H₂O₂ by the UV illumination (Eq. (7)), especially UV electromagnetic spectrum between 200 and 280 nm [4].



On the other hand, H₂O₂ can react with OH• and h⁺ to form the hydroperoxyl radical (HO₂•), especially when the concentration of H₂O₂ is high (Eqs. (8) and (9)), which can reduce the concentration of OH• (5).



The TiO₂ may be used either as an aqueous slurry or be immobilized onto a supporting substrate. When titania suspensions are used, the catalyst must be recovered at the end of the treatment, either by filtration or sedimentation which is expensive in terms

* Corresponding author. Tel.: +86 510 85918309; fax: +86 510 85918309.
E-mail address: huazz@jiangnan.edu.cn (Z. Hua).

of time, reagents and manpower. Alternatively, the TiO_2 can be immobilized onto a supporting substrate such as glass to avoid catalyst-recovering step [6]. Among the immobilized reactors, the immobilized TiO_2 thin-film reactors appear to be the most promising in terms of practical applications [7]. A thin-film catalyst can be prepared using a sol-gel technique and the supporting substrate coated with thin-film TiO_2 can be mounted in a reactor or be positioned outside of UV lamp to receive UV irradiation for the inactivation of cell.

It is clear that an in-depth understanding of the reaction mechanism responsible for photocatalytic inactivation of cells is helpful to increase the efficiency of photocatalytic disinfection system and devise a strategy in a practical system to efficiently kill microorganisms. However, there still exists controversy regarding the major species which is responsible for the photocatalytic bactericidal effect. It is frequently assumed that OH^\bullet are the main oxidative species responsible for the photocatalytic inactivation of bacteria [8–10]. Ireland et al. [8] reported that no inactivation of *E. coli* in the presence of the OH^\bullet scavenger was observed. The results of Sjogren and Sierka [9] showed that a 200% enhancement of its inactivation in the presence of a low concentration of ferrous ions. This observation of enhanced MS-2 phage inactivation was explained by the increased OH^\bullet concentration resulting from the Fenton reaction, as well as from the photocatalytic reaction ($\text{Fe}^{2+} + \text{H}_2\text{O}_2 \rightarrow \text{Fe}^{3+} + \text{OH}^- + \text{OH}^\bullet$, $\text{Fe}^{3+} + \text{e}^- \rightarrow \text{Fe}^{2+}$). However, there is also some evidence that H_2O_2 contribute to the photocatalytic inactivation of cells. For example, Kikuchi et al. [11] proposed that the main bactericidal source is not the OH^\bullet , but H_2O_2 in *E. coli* suspension separated from the TiO_2 thin film, since catalase that can decompose H_2O_2 greatly hindered the inactivation of *E. coli*, whereas the OH^\bullet scavenger did not. Maness et al. [12] reported that H_2O_2 and O_2^- , as well as the OH^\bullet , play significant role in microorganism inactivation. In these studies, the authors generally applied ROS scavengers such as mannitol and catalase or catalysts such as Fe^{2+} to validate their assumption, but seldom check the formations of ROS, especially the OH^\bullet . Obviously, monitoring ROS generation in photocatalytic reaction can provide direct evidence for identifying the primary bactericidal ROS. ESR spin-trapping technique appears to be one of the promising approaches for the detection of radicals' production, which has been used to observe directly ROS generate in TiO_2 photocatalytic reactor and provides essential information for understanding of the reaction mechanism on semiconductor surfaces [13,14]. In this work, to evaluate the bactericidal roles of H_2O_2 and OH^\bullet in immobilized TiO_2 thin-film reactor, the ESR spin-trapping technique using 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) as spin trap was applied to investigate the reaction mechanism of H_2O_2 and OH^\bullet in photocatalytic reaction. The bacte-

ricidal experiments were further performed using *Bacillus* sp. F26 as a model bacterium. The effects of O_2 and H_2O_2 on photocatalytic bactericidal efficiency were investigated.

2. Materials and methods

2.1. Materials

The photocatalytic reactor used in the experiment was kindly provided by Professor Fumihide Shiraishi of Kyushu Institute of Technology (Japan). Photocatalyst was prepared according to the method reported by Shiraishi et al. [15] as follows: amorphous TiO_2 , prepared from titanium isopropoxide, was dissolved into an aqueous solution of H_2O_2 , which give a transparent yellow solution. After this solution was swabbed on the surface of a quartz glass tube of 140 mm long and 28 mm inside diameter, the glass tube was fired at 500°C for 1 h, by which a transparent thin film of TiO_2 on quartz glass tube was formed. The spin trap reagent DMPO was purchased from Sigma (USA) and stored at -20°C in a freezer. All solutions and reagents were prepared in deionized/distilled water, analytical reagent-grade chemicals were used (Aldrich, USA). All glassware used in these experiments was washed with distilled water, and then autoclaved at 121°C for 15 min.

2.2. Experimental procedures

The reactions were carried out in an annular-flow photocatalytic reactor as illustrated in Fig. 1(A). The photocatalytic reactor was a cylindrical plastic vessel in which a 6.0 W UV lamp (254 nm), protected by a quartz glass tube from direct contact with the aqueous solution, was located at the center. Another big quartz glass tube with a membrane of TiO_2 on its outside wall was used as a sheath of the quartz glass. ROS were generated when the UV lamp was on and solution flowed through the space between the inner wall of the vessel and the glass sheath.

The photocatalytic reactor, a pyrex glass bottle (Vessel), a cooling apparatus, a flow meter and a peristaltic pump were connected in a loop, showed in Fig. 1(B). 350 mL deionized water was poured into the mixed-flow vessel (800 mL) and then recycled by means of a peristaltic pump at a flow rate of 300 mL/min. The reaction was started by switching the UV lamp on. At the same time, aseptic air was sparged into the pyrex glass bottle at the rate of 6 L/min. Operational conditions for all runs were kept at pH 7.0, temperature at 28.0°C through a cooling apparatus. During the reaction, liquid sample was withdrawn at appropriate time intervals from the bottle and analyzed immediately by means of H_2O_2 quantification

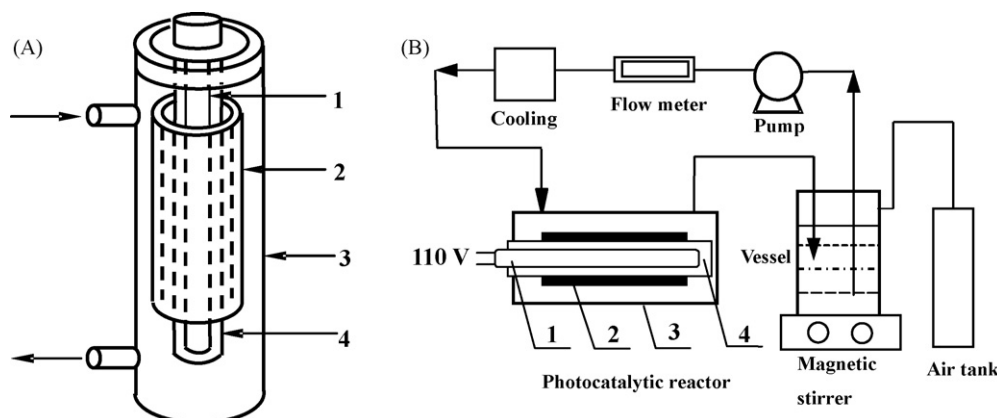


Fig. 1. Annular-flow photocatalytic reactor (A) and schematic diagram of experimental apparatus (B). (1) Ultraviolet lamp (6 W); (2) glass tube with photocatalyst membrane; (3) cylindrical plastic vessel (height: 20 cm, diameter: 5 cm); (4) quartz glass.

and ESR analysis described below. In the confirmation of OH^\bullet and O_2^- , dimethyl sulphoxide (DMSO), superoxide dismutase (SOD) and catalase (CAT) were introduced into the pyrex glass bottle prior to UV illumination, respectively.

2.3. Analytical techniques

The ESR spectrometer was used to detect and monitor the free radicals generated in reactor. The spin trap reagent, DMPO was dissolved in deionized water under the deair condition full of nitrogen and separated in 1 mL cone test tubes before experiments. In order to improve the noisy baseline ESR spectrum, the deionized water was sparged with helium gas for 2 h to remove any oxygen dissolved in the water before adding DMPO. The 0.5 mL sample contained ROS taken from the mixed-flow vessel was introduced into the cone test tube and mixed with the DMPO solution for reaction. The final reaction volume was 0.8 mL and the concentration of DMPO is 0.5 mM. The sample (100 μL) was immediately transferred into quartz capillaries specially designed for ESR analysis after 30 s for reaction and ESR spectra were recorded with a Bruker EMX 10/12 spectrometer at room temperature. ESR measurements were conducted under the following conditions: centre field, 3480 G; sweep width, 200 G; microwave frequency, 9.766 GHz; microwave power, 20.0 mW; modulation frequency, 100 KHz; modulation amplitude, 3.0 G; scan, 84 s; time constant, 41.0 ms. number of scans, 5.

The H_2O_2 concentrations produced in photocatalytic reactor were determined according to the enzymatic method described by Shiraishi et al. [16].

2.4. Disinfection experiments

In order to evaluate bactericidal effects of H_2O_2 and OH^\bullet , *Bacillus* sp. F26, isolated from Haoji Soda Lake located in the Hulunbeir area of Inner Mongolia Autonomous Region (China) was used as the model microorganism [17]. Our previous study showed that the strain is highly resistant to the oxidative stress [18]. The photokilling experiments were conducted as follows: *Bacillus* sp. F26 was cultivated aerobically at 37 °C with continuous shaking at 200 rpm in modified Horikoshi I medium containing (per liter of distilled water) 10 g of glucose, 10 g of polypeptone (BBI), 5 g of yeast extract (Oxoid), 1 g of KH_2PO_4 , 0.2 g of MgCl_2 , 50 g of NaCl, and 10 g of Na_2CO_3 . To ensure synchronous growth, 20 h-old culture was inoculated into the fresh medium and cultivated for 16 h to its exponential phase. The cells were harvested at this stage and used in the experiments. Bacterial cells were collected by centrifugation at 500 \times g for 10 min at 4 °C and the bacterial pellet was washed three times with deionized water. The cells were re-suspended and diluted to 10^7 – 10^8 colony forming units (CFU)/mL with sterilized water. Cell suspensions were diluted with sterilized water in the pyrex glass bottle to the required cell density corresponding to 10^5 – 10^6 CFU/mL. The experiment subsequently began when the UV lamp was switched on and performed for 100 min at room temperature. Samples were taken at 20 min interval, diluted in 50 mM potassium phosphate buffer and plated on the cultured medium agar plates in triplicate. The number of colonies appearing on the plates after 24 h of incubation at 37 °C was counted in terms of CFU.

3. Results and discussion

3.1. Generation of H_2O_2 in TiO_2/UV thin-film photocatalytic reactor

Fig. 2 illustrates the time-course of H_2O_2 produced in photocatalytic reactor. It could be seen that the concentration of H_2O_2 increased with the illuminated time prolonged and the maximum value (0.033 mM) appeared after 12 h of illumination, but further

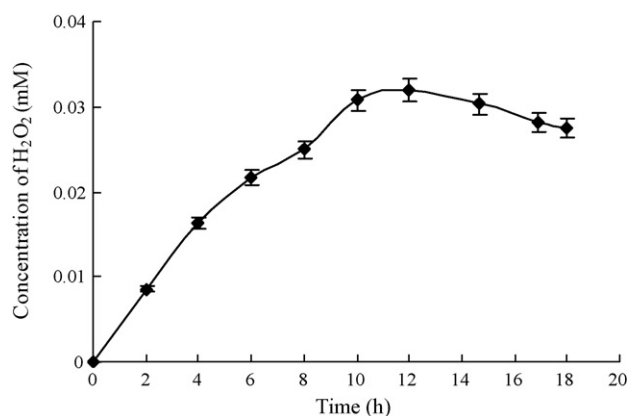


Fig. 2. Time-course of H_2O_2 concentration in TiO_2 thin-film reactor. Error bars indicate the S.D.

irradiation resulted in low concentration. According to Eqs. (1)–(4), H_2O_2 is mainly generated from the reduction reaction of O_2 (Eq. (4)). Once formed, it would convert to OH^\bullet through the further reduction (Eqs. (5) and (6)) and the photolysis (Eq. (7)). Meanwhile, H_2O_2 can also react with OH^\bullet and h^+ to form the HO_2^- (Eqs. (8) and (9)), which could result in reduction of H_2O_2 concentration. However, according to the results of Bekbolet and Balcioglu [5] that the oxidation reaction of H_2O_2 was minor compared to the reduction reaction unless higher concentrations of H_2O_2 (above 10 mM) were present, in our case, it was believed that the contribution of oxidation reaction to H_2O_2 level decline was insignificant due to its low concentration. Our results suggested that the H_2O_2 generation from O_2 reduction was faster than H_2O_2 decomposition (Eqs. (5)–(7)) before 12 h of irradiation. After that, the latter exceeded the former.

3.2. ESR measurement of ROS generated in TiO_2/UV thin-film photocatalytic reactor

Fig. 3 presents the time-course of ESR spectra measured in photocatalytic reactor in the presence of DMPO as the spin trap. It could be observed that the intensities of signal strengthened with increased irradiation time, indicating the concentrations of free radicals increased with the irradiation time prolonged. The ESR spectrums are composed of a four-line signal, having a peak height ratio of 1:2:2:1 signal pattern and hyperfine coupling constant ($\alpha_N = \alpha_H = 14.9$ G) matched with those of typical DMPO–OH adduct formed upon trapping of OH^\bullet by the DMPO [19].

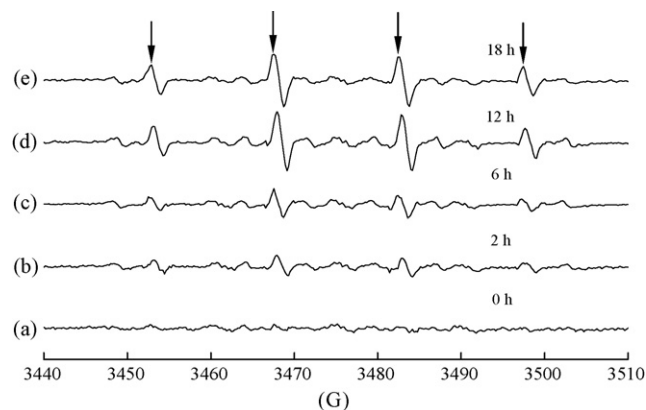


Fig. 3. ESR spectra of DMPO adducts measured in TiO_2 thin-film reactor. Spectra a–e denote the irradiation time 0, 2, 6, 12 and 18 h, respectively. (Arrow denotes the position of hyperfine peaks of DMPO–OH adducts).

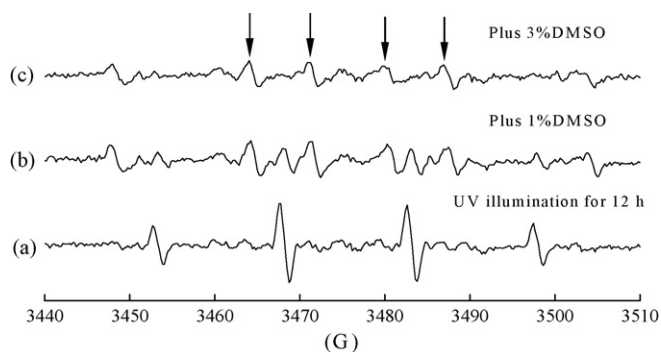


Fig. 4. ESR spectra of the DMPO adducts in 1% and 3% DMSO. (a) After 12 h of irradiation; (b) same as (a), except that 1% DMSO was added into the reactor prior to irradiation; (c) same as (a), except that 3% DMSO was added into the reactor prior to irradiation. (Arrow denotes the position of hyperfine peaks of DMPO-CH₃ adducts).

It should be pointed out that the observed increase in the DMPO-OH signals may be due to the decay of the O₂⁻ adduct, DMPO-OOH, or from a combination of both process. Therefore, 1% and 3% DMSO were pre-added into the reactor (Fig. 4). DMSO can react with OH• to give CH₃ radicals which can react with DMPO to yield DMPO-CH₃ with its characteristic hyperfine splitting pattern. The results showed that the signal of DMPO-OH decreased and disappeared in the presence of 1% and 3% DMSO, respectively, but presented features corresponding to the DMPO-CH₃ adduct ($\alpha_N = 16.4$ G, $\alpha_H = 23.3$ G) [20]. This suggests that the DMPO-OH signal observed in the absence of DMSO originated from the trapping of OH• and not from the decomposition of DMPO-OOH. Further evidence for the source of the DMPO-OH signal was obtained by carrying out the reaction in the presence of SOD (40 μ g/mL), which can catalytically convert the O₂⁻ to H₂O₂ and O₂. It was found that the signal of OH• adducts were insensitive to SOD and inactivated SOD (date not shown). The similar phenomena were observed in the samples after illuminated for 2, 6 and 18 h (date not shown), suggesting that the intensities of ESR spectrums in Fig. 3 were induced by OH• adducts.

Although O₂⁻ should be formed as a result of the scavenging of the electron by O₂, we were unable to detect directly the spin-adduct of this radical under the present condition. Similar results were reported by other authors [21–23]. This may be due to the fact that O₂⁻ in water is very unstable and undergo facile disproportionation rather than slow reaction with DMPO [21]. Alternatively, it can be explained by the assumption that O₂⁻ is not desorbed or that it reacts quickly towards other products (e.g., H₂O₂) [23]. In fact, the

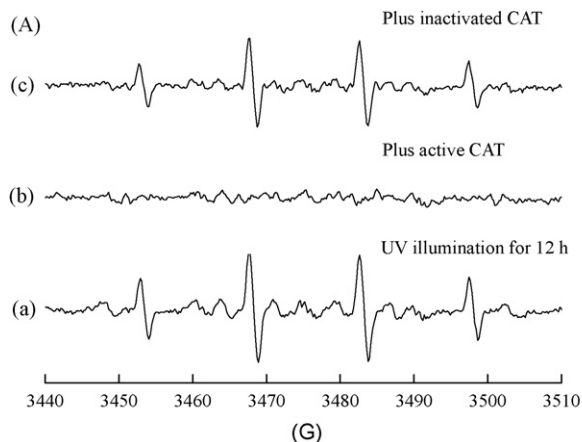


Fig. 6. ESR spectra of the DMPO adduct in CAT (A) and exogenous H₂O₂ without and with TiO₂ (B). (a) After 12 h of irradiation; (b) same as (a), except that active CAT (4 mg/mL) was added prior to irradiation; (c) same as (a), except that inactivated CAT (by heat) was added prior to irradiation; (d) 1 mM H₂O₂ was irradiated for 2 h in the absence of TiO₂; (e) 1 mM H₂O₂ was irradiated for 2 h in the presence of TiO₂.

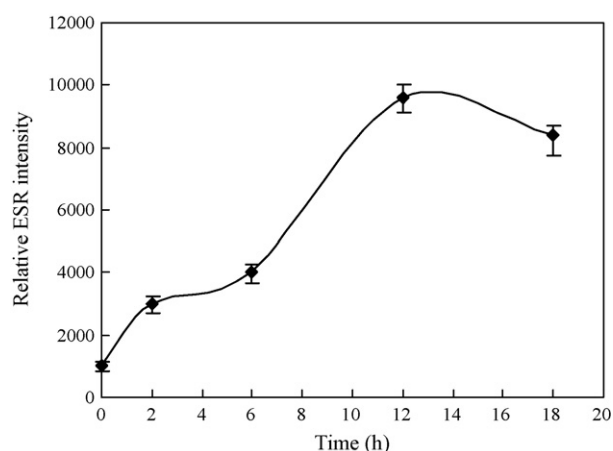


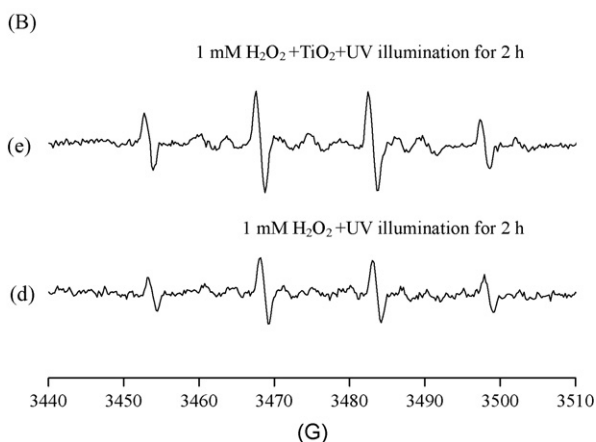
Fig. 5. Evolution of the DMPO-OH signal intensity (evaluated from the amplitude of the second line of the signal) as a function of the time of irradiation. Error bars indicate the S.D.

role of O₂⁻ in photocatalytic disinfection was demonstrated to be insignificant relative to H₂O₂ and OH•. For example, the results of Cai et al. showed that antibacterial efficiency greatly increased in the presence of SOD and Fenton reagent due to the formation of H₂O₂ and OH• from O₂⁻ [24].

To determine the relative levels of OH• formed in photocatalytic reaction, evolution of the DMPO-OH• signal intensity as a function of irradiation time is presented in Fig. 5, which were evaluated from the amplitude of the second line of the signal [25]. It showed that like the profile of H₂O₂, the intensity of DMPO-OH adduct gradually enhanced with illumination time increased and reached the peak at 12 h, after that, began to decline. Similar results were reported by Brezova et al. [26]. These results suggested that the production of H₂O₂ can promote the generation of OH•. With regard to the decrease of DMPO-OH• signal intensity by prolong irradiation, it might be due to the multiple addition of OH• to DMPO, producing ESR-silent products [26].

3.3. The transformation of H₂O₂ into OH•

Numerous studies have demonstrated that there is a good correlation between the photocatalytic bactericidal efficiency and the amount of OH•. Therefore, it is important to clarify the originating source of OH• in photocatalytic reaction to improve the efficiency of photocatalytic sterilization. According to Eqs. (1)–(7), OH• are



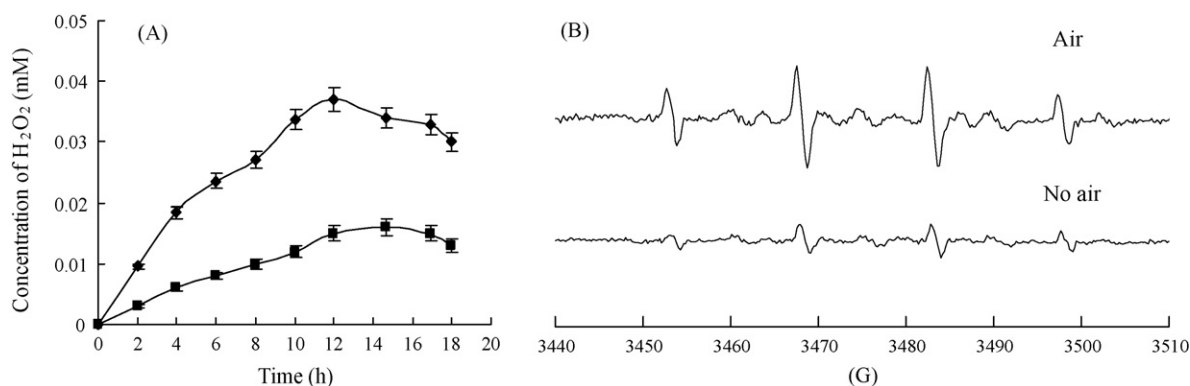


Fig. 7. Comparisons of H₂O₂ concentration (A) and ESR spectra (B) with and without air. (◆) 10 L/min of air was introduced into the vessel in the presence of UV illumination and TiO₂; (■) no air was introduced into the vessel. The ESR spectra of DMPO adduct were obtained after 12 h illumination. Error bars indicate the S.D.

produced from the following three routes: (1) reaction of positive hole (h⁺) with H₂O (Eq. (2)); (2) the reduction reaction of H₂O₂ with electron (Eqs. (5) and (6)) and (3) direct decomposition of H₂O₂ by UV light (Eq. (7)), especially UV spectrum is located between 200 and 280 nm [4]. To elucidate which pathway among pathway 1–3 is the main route to the OH[•] formation, ESR experiments were performed in the presence of catalase (CAT), which catalytically converts the H₂O₂ to H₂O and O₂. As showed in Fig. 6A, the signals of DMPO–OH adduct disappeared after addition of active CAT (4 mg/mL), but the inactivated CAT did not produce similar results. Obviously, H₂O₂ is indispensable for the formation of OH[•]. OH[•] may either come from the H₂O₂ reduction or from direct photolysis, or a combination of both pathways. The ESR experiment was further carried out to identify which is the primary route. To strengthen the signal of ESR spectra for easy comparison, 1 mM H₂O₂ was added into the reactor with TiO₂ (H₂O₂/UV/TiO₂) and without TiO₂ (UV/H₂O₂) prior to UV illumination, respectively. The discrepancies in the ESR spectra were compared after 2 h of irradiation (Fig. 6B). In addition, the remained H₂O₂ concentration was measured.

It was obvious that the intensities of OH[•] adducts signal in UV/H₂O₂ decreased when TiO₂ film was removed. With regard to H₂O₂, its level dropped by about 28.1% in UV/H₂O₂ over the 120 min experiment time. In contrast, its level in the H₂O₂/UV/TiO₂ declined much more steeply with a 76.3% decrease after 120 min. These results indicated that TiO₂ could speed up the conversion of H₂O₂ to OH[•] and OH[•] were mainly originated from the H₂O₂ reduction. Previous studies showed that addition of H₂O₂ could greatly increase the efficiency of UV disinfection as a result of elevation of OH[•] by H₂O₂ direct photolysis; however, it was proved that its inactivated efficiency was lower compared with H₂O₂/UV/TiO₂ [27]. Our results well supported this conclusion.

Previous studies have showed that the introduction of pure oxygen or air can enhance the generation of H₂O₂ [28]. To evaluate the effect of endogenous H₂O₂ on the formation of OH[•], the influences of air bubble (10 L/min and void air) on the formations of H₂O₂ and ESR spectra were investigated. As expected, the rate of H₂O₂ formed was accelerated by bubbling 10 L/min air and accordingly the maximum concentration increased by 131.2% when compared to those without air (Fig. 7A), the highest value of H₂O₂ was also 16.7% higher than the value obtained in 6 L/min (Fig. 2). Similarly, the intensity of DMPO adducts was enhanced by air bubble relative to that without air (Fig. 7B). This suggested that increased O₂ provided by air bubbles react with the electron trapped on the surface of TiO₂ and generate O₂^{•-}. It further reacts with electron and form H₂O₂ (Eq. (4)), which then decomposed and produces OH[•] [3].

It is well known that the formation and transformation of oxygen radicals in photocatalytic reaction are dependent on their potential for oxygen reduction. Due to the truth that the potential for O₂ reduction is –0.13 V while that of H₂O₂ reduction is 0.72 V, H₂O₂ is

a better electron acceptor than oxygen. This means that once H₂O₂ appear, the most electrons excited from band will turn to H₂O₂ for producing OH[•] instead of reacting with O₂ to form O₂^{•-}. As a result, the accumulation of H₂O₂ was always accompanied by increase of OH[•] (Figs. 2 and 5). On the other hand, the failure of detecting O₂^{•-} in this study could be explained by the assumption that the amount of O₂^{•-} in reactor could not reach the level that efficiently trapped by DMPO due to the presence of H₂O₂.

In our study, it was found that the signal of DMPO–OH adduct was not immediately disappear after turn-off of the UV light, instead, it decayed much slowly and could be still observed after 20 min in the dark, implying that the photocatalytic reaction can continue to take place in the dark and lead to the OH[•] formation (Fig. 8). The similar results were reported by Liu et al. [22]. In photocatalytic inactivation study, it was also found that cells continue to lose their viability even after stop UV irradiation, which was called residual disinfection [29]. Maness et al. attributed it to the assumption that certain lethal reaction can continue to propagate after the UV illumination stops [12]. Another explanation is that oxidative species continue to generate and attack cellular membrane and probably enter into the cells leading to the bacterial cell death [30]. Our results in Fig. 8 seem to support the second assumption. Since the addition of CAT (4 mg/mL) resulted in disappearance of DMPO–OH adducts' signal after UV light was turned off (date not shown), it is assumed that OH[•] may come from the decomposition of H₂O₂ formed in the photocatalytic reaction. The detailed mechanism need to be further investigated.

3.4. The important roles of H₂O₂ and OH[•] in the bactericidal effects of TiO₂/UV thin-film photocatalytic reactor

Our results showed that the levels of OH[•] and H₂O₂ synchronously enhanced with the illumination time increased as a result of conversion of H₂O₂ to OH[•] (Figs. 2 and 5). In order to

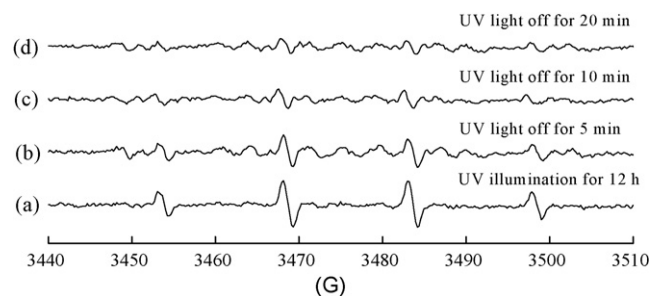


Fig. 8. Changes of DMPO adduct ESR spectra after UV light off. (a) After 12 h of irradiation; spectra b–d denote after 5, 10 and 20 min of UV light off, respectively.

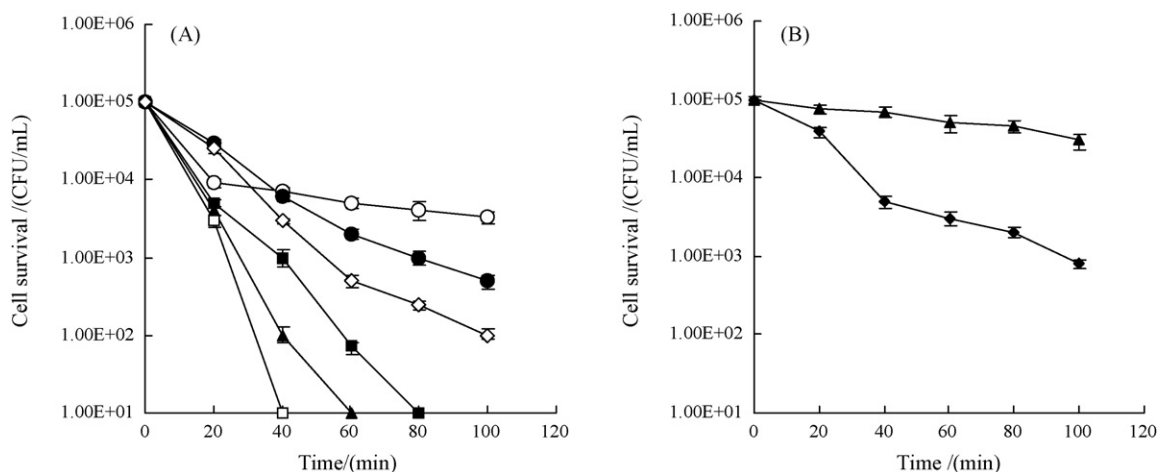


Fig. 9. Effects of air bubble and H₂O₂ (A) and catalase (B) on bactericidal efficiency of TiO₂ thin-film reactor. A: ● Control: 6 L/min of air was introduced into the vessel in the presence of UV illumination and TiO₂; ◇ Same as control, except that 10 L/min of air was introduced; ○ 6 mM H₂O₂ was added into the vessel without UV illumination; ■ Same as control, except that 1 mM H₂O₂ was added into the vessel prior to irradiation; ▲ Same as control, except that 3 mM H₂O₂ was pre-added; □ Same as control, except that 6 mM H₂O₂ was pre-added. B: ◆ Control: 6 L/min of air was introduced into the vessel in the presence of UV illumination and TiO₂; ▲ Same as control, except that CAT (4 mg/mL) was added into the vessel prior to irradiation. Error bars indicate the S.D.

determine their roles in killing effect, the bactericidal experiments were carried out using *Bacillus* sp. F26 as a mode microorganism. Considering the fact that UV lamp of 254 nm have germicidal effect, the control experiments were firstly conducted, in which TiO₂ film or UV light was removed from the photocatalytic reactor, respectively. The results showed that no obvious changes in bacteria survival in the absence of TiO₂ film or UV light, suggesting that disinfections without photocatalyst or in the dark were insignificant (data not shown).

Fig. 9A showed that the influences of bubbling air and H₂O₂ addition on photocatalytic killing effects. The bactericidal efficiency was enhanced by around 5-fold within 100 min of UV illumination in the 10 L/min air compared to the control (6 L/min), suggesting that increased H₂O₂ and OH• levels was induced by providing extra O₂. In contrast, increasing H₂O₂ concentration from 1 to 3 and finally to 6 mM resulted in a drastic inactivation effect, no viable cells were found after 40, 60 and 80 min irradiation in the presence of 6, 3, 1 mM H₂O₂. This suggests that a great amount of OH• was generated with the elevation of H₂O₂ through decomposition, and also indicates that H₂O₂ was more efficient than O₂ to increase bactericidal effect in TiO₂ film photocatalytic reactor due to its higher capability to react with electron [31]. It should be noted that H₂O₂ (6 mM) alone has the capability of killing cells. The survival number of cells decreased from 1 × 10⁵ to 1 × 10⁴ CFU/mL within 20 min, then after, the killing effect was negligible. This may be explained by the fact that H₂O₂ induced catalase production within *Bacillus* sp. F26 and therefore decomposed the remained H₂O₂ [18]. The above results indicated that OH•, not H₂O₂ was the major bactericidal radical in TiO₂ thin-film reactor. However, considering the convertible relationship between H₂O₂ and OH•, H₂O₂ was indispensable for photocatalytic bactericidal effect. To verify this assumption, H₂O₂ scavenger, CAT was added prior to illumination. As expected, no significant bactericidal effect was observed in the presence of 4 mg/mL CAT (Fig. 9B). This result also validated the conclusion from the control experiment that the germicidal effect of UV light alone was negligible, and the inactivation of *Bacillus* sp. F26 in TiO₂ thin-film reactor was attributed to ROS produced in the photocatalytic reaction.

Besides providing OH•, the possibility that H₂O₂ directly induce cells damage could not be precluded. It was proposed that the photokilling process consists of two steps [32]. The cell wall was initially attacked by OH•, followed by a progressive damage of the cytoplasmic membrane and intracellular components caused by other ROS

attack, such as H₂O₂. Therefore, once the cells wall was broken or antioxidative systems were disrupted, H₂O₂ would easily enter into the cells to accelerate the cells death. In addition, when the membrane was perforated, Fe²⁺ and Cu²⁺ could be freely released into the environment and react with H₂O₂ to form OH•, result in aggravation of cell damage [3]. Our results in Fig. 9A validated the hypothesis, in that no discrepancy in cells number before 20 min in the presence of 1, 3, 6 mM H₂O₂ was observed, whereas, the rates of cell death were speeded up after 20 min with the H₂O₂ concentration elevated. On the basis of these results, it can be concluded that it is a cooperative work of OH• and H₂O₂ to complete the bactericidal process, in which OH• was the primary bactericidal oxygen radicals, and H₂O₂ as a supplier of OH• and another important attack radicals was also indispensable for disinfection effect.

4. Conclusions

In this study, the mechanism of H₂O₂ and OH• formation and transformation in the immobilized TiO₂ thin-film reactor was investigated by applying ESR spin-trapping technique. The results showed that the level of OH• was well associated with the amount of H₂O₂ because OH• was mainly originated from the H₂O₂ via the reduction reaction although a relatively small portion came from the direct photolysis of H₂O₂. The bactericidal efficiency of reactor was proportionally increased with elevated H₂O₂ levels, whereas, H₂O₂ alone was not enough to complete the disinfection action. Based on these results, it is concluded that OH•, not H₂O₂ was the primary bactericidal radicals in TiO₂ thin-film reactor; however, due to its conversion to OH• and oxidative toxicity, H₂O₂ was also indispensable for photocatalytic bactericidal effect.

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